

Antioxidant activity of *Mitragyna parvifolia* (Roxb.) Korth

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Abstract

Antioxidants play a crucial role in protecting living organisms against oxidative stress, a condition associated with various chronic diseases and aging. Extensive research has been conducted to identify and to characterize natural compounds with potent antioxidant activity, aiming to harness their potential benefits for human health. Mitragyna parvifolia (Roxb.) Korth, a member of the Rubiaceae family, has gained considerable importance in the field of traditional medicine, particularly in the system of Ayurveda. This plant has been extensively utilized for various purposes, solidifying its significance in traditional medicinal practices.

This research aims to investigate the antioxidant activity of Mitragyna parvifolia using different methods. The plant extract was evaluated by 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging assay, ferric reducing antioxidant power (FRAP) assay, nitric oxide free radical scavenging activity assay and phosphomolybdate assay. The results provide valuable insights into the potential applications in the development of natural antioxidants from Mitragyna parvifolia.

Keywords: *Mitragyna parvifolia*, Rubiaceae, Antioxidant activity, DPPH, FRAP, Nitric oxide, Phosphomolybdate.

Introduction

Our bodies engage in various natural biological processes like breathing, digesting food and converting fats into energy. However, these processes also generate harmful substances known as free radicals. Normally, our body's natural antioxidant system eliminates these free radicals. But when this system fails to function properly, free radicals can initiate harmful reactions within the body. This reaction has the potential to damage cell membranes, to hinder the activity of crucial enzymes, to disrupt essential cellular processes, to impede normal cell division, harm DNA and to obstruct energy production¹⁷. Free radicals have a vital role in numerous biological processes, some of which are crucial for sustaining life.

Free radicals, they play a role in the intracellular eradication of bacteria by phagocytes, notably granulocytes and macrophages. Additionally, researchers have identified the involvement of free radicals in specific cellular signaling

processes, commonly referred to as redox signaling⁵. The interaction between free radicals and DNA is believed to contribute to the development of numerous cancers. This interaction can cause mutations that disrupt the normal cell cycle, ultimately leading to the formation of neoplasms²³. Free radicals and reactive oxygen species are integral components of life, engaging in crucial biological reactions²⁵. The presence of antioxidant compounds is necessary to neutralize the harmful impacts of reactive oxygen species²⁴.

Chronic diseases such as cancers, arthritis, cardiovascular disease and vascular disorders are primarily impacted by oxidative stress. Plants, with their diverse range of antioxidant compounds, serve as a valuable reservoir of active biological substances²⁵. Antioxidants play a protective role against oxidants by either eliminating free radicals or inhibiting their formation thus safeguarding cells from damage⁶. Phenolic compounds exhibit antioxidant activity by effectively neutralizing free radicals²⁵. Plant phenolic compounds perform a variety of functions, such as having antioxidant properties, exhibiting antibacterial and natural pesticide properties, functioning as plant protection agents against UV waves and serving as insulating agents for plant cell walls against gases²¹.

Phenolic compounds are known as excellent sources of natural antioxidants within plants. Among them, flavonoids represent the largest group of plant phenols, accounting for over half of the approximately 8,000 natural phenolic compounds²⁰. The antioxidant potency of flavonoid compounds is directly linked to the increase in the number of hydroxyl groups they possess¹⁶. Plants have historically served as a valuable source of exogenous antioxidants for dietary purposes. It is estimated that approximately two-thirds of the world's plant species hold medicinal significance, with nearly all of them possessing remarkable antioxidant potential¹². Medicinal plants, also known as medicinal herbs, have been utilized in traditional medicine for centuries. These plants produce a wide array of chemical compounds that serve various purposes, such as defense against insects, fungi and diseases³.

Rubiaceae, a plant family of significant medicinal and economic importance, is found across various regions. The plant members belonging to this family holds immense medicinal value due to the diverse range of phytochemicals present within them⁴. *Mitragyna parvifolia*, commonly known as Kadamb and belonging to the Rubiaceae family, is a tree that holds substantial cultural, health and economic significance⁸. It is found in deciduous and evergreen forests

across India, contains various chemical constituents, including pyroligneous acid, methyl acetate, ketones and aldehydes. Because of its numerous medicinal properties, the plant is extensively utilized by tribal communities and Ayurvedic practitioners¹³.

Extensive scientific research has established that this medicinal plant exhibits a wide range of immunopharmacological properties including analgesic, anti-pyretic, anti-inflammatory, anti-arthritis, anti-helminthic and antioxidant effects⁷. The bark and roots of the plant are utilized in the treatment of various conditions such as fever, colic, muscular pain, burning sensation, poisoning, gynecological disorders, cough and edema. Additionally, fruit juice is known to enhance breast milk production in lactating mothers¹⁵. The objective of this study is to assess the antioxidant activity of leaves of *Mitragyna parvifolia* by employing multiple assay methods.

Material and Methods

Mitragyna parvifolia leaves were collected from Vile parle and authenticated at Blatter Herbarium, St. Xaviers College, Mumbai (Accession no. 433). The leaves were washed, air dried and ground into a fine powder using blender. The aqueous and hydro alcoholic leaf extract was prepared using Soxhlet apparatus. The scavenging ability of *Mitragyna parvifolia* leaf extract against DPPH (2,2-diphenyl-1-picrylhydrazyl) free radicals was evaluated using a standard method with appropriate modifications according to the method of Kaskoos¹¹. The reducing power of *Mitragyna parvifolia* leaf extract was determined according to the method of Narayanan et al¹⁸ whereas nitric oxide free radical scavenging activity was determined according to the method of Olatidoye et al¹⁹. The total antioxidant capacity assay was conducted using the phosphomolybdenum method⁹.

Results and Discussion

Free radical scavenging activity: DPPH is a free radical compound commonly employed to assess the free radical-scavenging potential of various samples¹⁰. This method involves measuring the ability of substances to act as hydrogen providers or free-radical scavengers by interacting with the stabilized free radical DPPH¹. Figure 1 shows the DPPH radical scavenging activity of aqueous (D/W) and hydro alcoholic (HA) plant extract compared to ascorbic acid (AA). As concentration increases, the antioxidant activity increases. The aqueous and hydro alcoholic leaf extracts show similar activity as compared to standard ascorbic acid.

Reducing power assay: The FRAP method relies on antioxidants capability to convert Fe^{3+} ions to Fe^{2+} , aligning the antioxidant strength with this reducing ability. The introduction of TCA solution causes the precipitation of the potassium ferrocyanide complex and the addition of FeCl_3 results in the formation of a green to blue complex known as Berlin blue. Measurement of the color complex at 700 nm allows the determination of the reduction ability, indicating the potential antioxidant activity of compounds. Compounds with strong reduction abilities are likely to function as antioxidants by stabilizing radicals through electron or hydrogen atom donation, enhancing the stability of radical compounds²².

Figure 2 shows the reductive capabilities of aqueous(D/W) and hydro alcoholic (HA) plant extracts compared to ascorbic acid (AA). Ferric ion reducing antioxidant power activity shows that as the concentration increases, the reducing power also seems to increase. However, the aqueous and hydro alcoholic leaf extracts show less activity as compared to standard ascorbic acid.

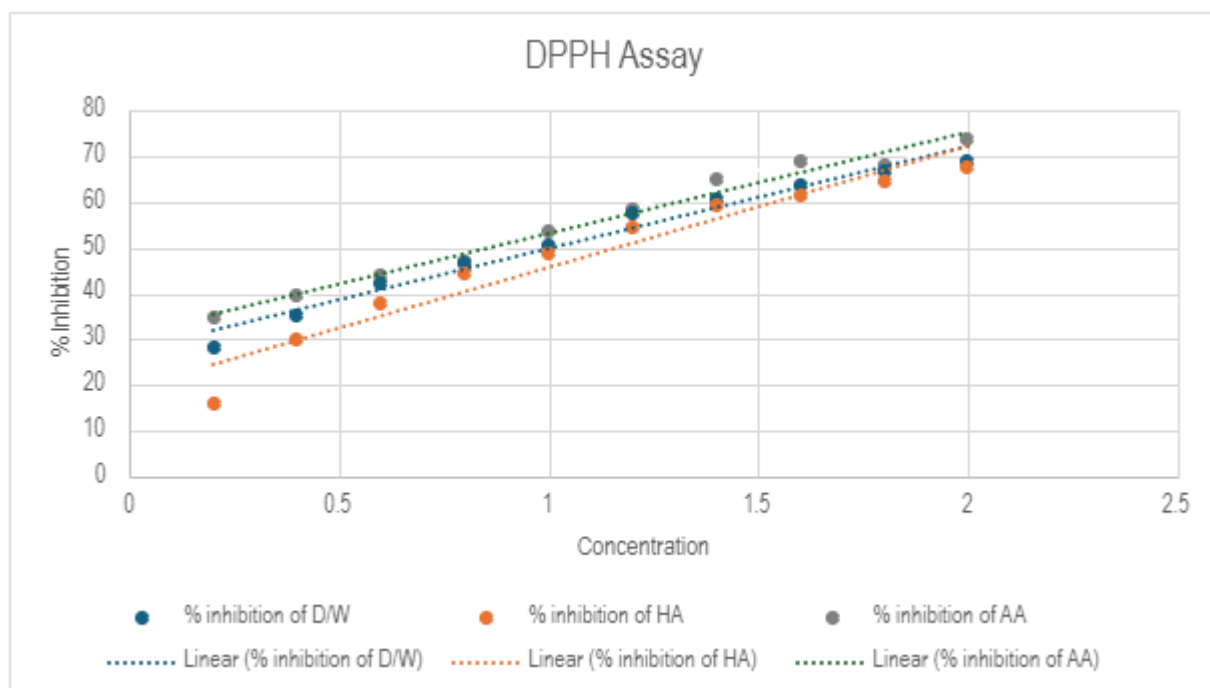


Figure 1: Free radical scavenging activity of *Mitragyna parvifolia* leaf extract

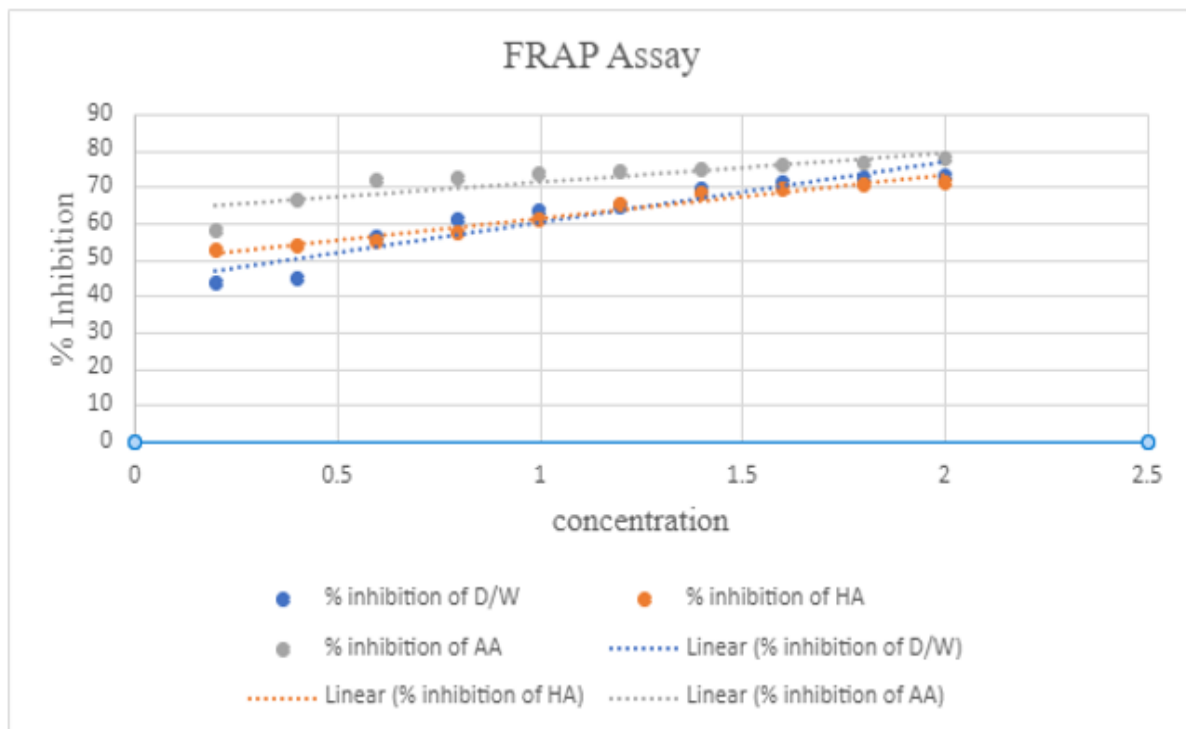


Figure 2: Reducing power assay of *Mitragnya parvifolia* leaf extract

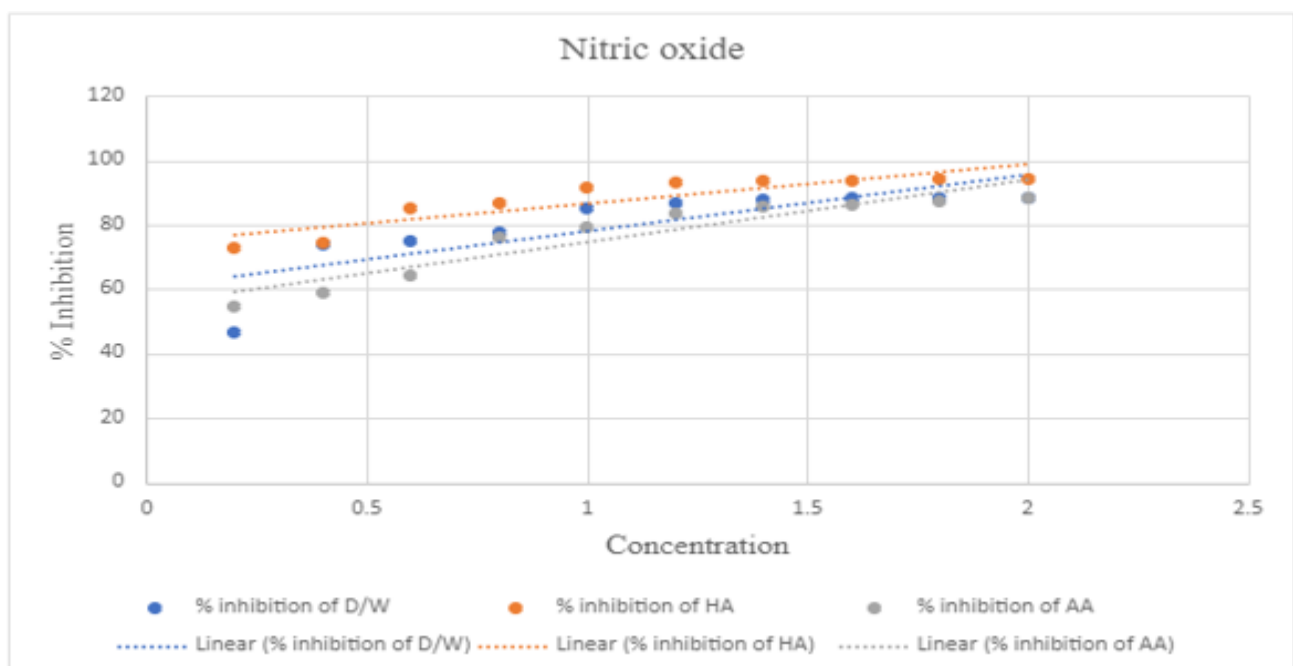


Figure 3: Nitric oxide scavenging activity of *Mitragnya parvifolia* leaf extract

Nitric oxide scavenging activity: Nitric oxide (NO) is a highly reactive diatomic free radical with a brief lifespan in biological systems. An indirect method to determine NO spectrophotometrically involves measuring its stable decomposition products NO_3 and NO_2 . This process entails reducing NO_3 to NO_2 followed by determining NO_2 , through the Griess reaction. The Griess reaction involves a two-step diazotization reaction where the nitrosating agent N_2O_3 reacts with sulfanilamide, forming a diazonium ion. This ion couples with N-(1-naphthyl) ethylenediamine, producing a chromophoric azo product with strong absorption at 540 nm.

Enzymatic reduction of NO_3 to NO_2 using a commercially available nitrate reductase has proven to be an effective method for quantifying these compounds in extracellular fluids². Figure 3 shows the nitric oxide scavenging activity of aqueous (D/W) and hydro alcoholic (HA) plant extract compared to ascorbic acid (AA). Nitric oxide scavenging activity shows that as the concentration increases, the activity goes on increasing rapidly, but as the concentration reaches $1\mu\text{g/ml}$ and above, the activity does not increase exponentially. However, the activity of aqueous and hydro alcoholic leaf extract is greater than standard ascorbic acid.

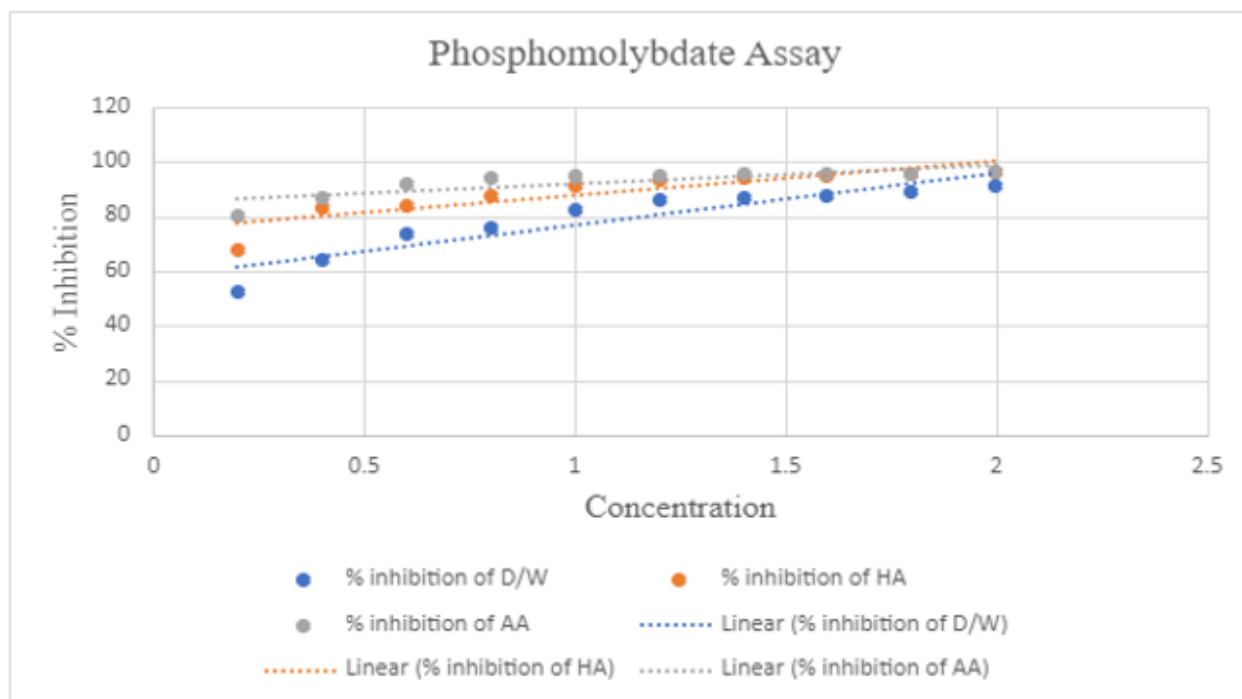


Figure 4: Phosphomolybdenum assay of *Mitragyna parvifolia* leaf extract

Phosphomolybdenum assay: This assay involves the extract reducing Mo (VI) to Mo (V), leading to the formation of a green phosphate/Mo (V) complex under acidic pH conditions¹⁴. Figure 4 shows the reducing Mo (VI) to Mo (V) activity of aqueous (D/W) and hydro alcoholic (HA) plant extract compared to ascorbic acid (AA). The percentage total antioxidant activity shows an increase in the activity as the concentration increases. In comparison to standard ascorbic acid and hydro alcoholic leaf extract, the aqueous leaf extract has lower activity. However, the hydro alcoholic leaf extract has comparable activity with standard ascorbic acid.

Conclusion

The study demonstrates that both aqueous (D/W) and hydro alcoholic (HA) plant extracts exhibit significant antioxidant activities as evidenced by their performance in DPPH, FRAP, nitric oxide scavenging and phosphomolybdenum reduction assays. The antioxidant activity of the plant extracts generally increases with concentration. The DPPH assay shows that both extracts have similar scavenging activity to ascorbic acid whereas the FRAP assay indicates a lower reducing power for the extracts compared to ascorbic acid.

The nitric oxide scavenging assay indicates that both extracts show greater scavenging activity than ascorbic acid at higher concentrations while the phosphomolybdenum assay shows that the hydro alcoholic extract has comparable activity to ascorbic acid and the aqueous extract exhibits lower activity. These results suggest that the hydro alcoholic extract, in particular, may serve as an effective antioxidant. The result of the antioxidant activity of leaves of *Mitragyna parvifolia* highlights its potential as a natural source of antioxidants. Further research is warranted to explore the practical

applications of *Mitragyna parvifolia* in preventing oxidative stress-related diseases and developing antioxidant-rich pharmaceutical or nutraceutical products.

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References

- Baliyan Siddhartha, Mukherjee Riya, Priyadarshani Anjali, Vibhuti Arpana, Gupta Archana, Pandey Ramendra Pati and Chang Chung-Ming, Determination of Antioxidants by DPPH Radical Scavenging activity and Quantitative Phytochemical analysis of *Ficus religiosa*, *Molecules*, **27**(4), 1326 (2022)
- Bryan Nathan S. and Grisham Matthew B., Methods to Detect Nitric oxide and its Metabolites in Biological samples, *Free Radic Biol Med*, **43**(5), 645-657 (2007)
- Chaudhari Amit and Ghogare Pradip, Review on Malvaceae and Rubiaceae Family, *Journal of Pharmacognosy and Phytochemistry*, **10**(4), 1-24 (2022)
- Das B.P. and Ghosh A., Medicinal properties of Phyto-constituents from Rubiaceae family of Assam: Critical Review, *International Journal of Applied Chemical and Biological Sciences*, **1**(3), 1-25 (2020)
- Finkel T. and Holbrook N., Oxidants, oxidative stress and the biology of ageing, *Nature*, **408**(6809), 239-247 (2000)
- Galhardo D., Garcia R.C., Schneider C.R., Braga G.C., Chambó E.D., França D.L.B. and Ströher S.M., Physicochemical, bioactive properties and antioxidant of *Apis mellifera* l honey from western Paraná, Southern Brazil, *Food Science and Technology*, **41**(1), 247-253 (2021)

7. Gupta Amit and Chaphalkar Sushama R., Immunosuppressive and Cytotoxic Potential of Flavonoids from *Mitragyna parvifolia*, *Mangifera indica* and *Aegle marmelos*, *Research & Reviews Journal of Pharmacology and Toxicological Studies*, **4**(1), 1-5 (2016)
8. Gupta Vikas, Kumar Pawan, Bansal Parveen and Singh Ranjit, Anti-inflammatory and Anti-nociceptive Activity of *Mitragyna parvifolia*, *Asian Journal of Medical Sciences*, **1**(3), 97-99 (2009)
9. Jan Shumaila, Khan Muhammad Rashid, Rashid Umbreen and Bokhari Jasia, Assessment of antioxidant potential, total phenolics and flavonoids of different solvent fractions of *Monothecha buxifolia* fruit, *Osong Public Health Res Perspect.*, **4**(5), 246-54 (2013)
10. Joshi Hardik, Shingadia Hitesh and Vaidya Meenakshi, In-vitro Antioxidant activity of Methanolic extract of leaves of *Lannea coromandelica* (Hout.)Merill, *J. Indian Bot.Soc.*, **91**(4), 408-413 (2012)
11. Kaskoos Raad A., Physico chemical parameters Phytochemical screening and Antioxidant activity of seeds of *Peganum harmala* collected from Iraq, *Asian Journal of Biomedical and Pharmaceutical Sciences*, **4**(28), 20-24 (2014)
12. Kasote D.M., Katyare S.S., Hegde M.V. and Bae H., Significance of antioxidant potential of plants and its relevance to therapeutic applications, *Int J Biol Sci.*, **11**(8), 982-91 (2015)
13. Kaushik D., Khokra S.L., Kaushik P., Saneja A., Sharma C., Aneja K.R., Chaudhary B. and Koshy S., A Study of Analgesic and Antimicrobial Potential of *Mitragyna Parvifolia*, *International Journal of Pharmaceutical Sciences and Drug Research*, **1**(1), 6-8 (2009)
14. Khatoon Mahfuza Islam Ekramul, Rafikul Islam, Rahman Abdur Aziz, Alam A.H.M. Khurshid, Khondkar Proma, Rashid Mamunur and Parvin Shahnaj, Estimation of Total phenol and in vitro antioxidant activity of *Albizia procera* leaves, *BMC Research Notes*, **6**, 121 (2013)
15. Kotval Shailesh C., Tessy John and Parmar Kokila A., Green synthesis of copper nanoparticles using *Mitragyna parvifolia* plant bark extract and its antimicrobial study, *J. Nanosci. Tech.*, **4**(4), 456-460 (2018)
16. Lou X., Xu H., Hanna M. and Yuan L., Identification and quantification of free, esterified, glycosylated and insoluble-bound phenolic compounds in hawthorn berry fruit (*Crataegus pinnatifida*) and antioxidant activity evaluation, *LWT*, **130**, 109643 (2020)
17. Mehdi Sharifi-Rad, Nanjangud V. Anil Kumar, Zucca Paolo, Varoni Elena Maria, Dini Luciana, Panzarini Elisa, Rajkovic Jovana, Tsouh Fokou Patrick Valere, Azzini Elena, Peluso Ilaria, Mishra Abhay Prakash, Nigam Manisha, Rayess Youssef El, Beyrouthy Marc El, Polito Letizia, Iriti Marcello, Martins Natalia, Martorell Miquel, Docea Anca Oana, Setzer William N., Calina Daniela, Cho William C. and Sharifi-Rad Javad, Lifestyle, Oxidative Stress and Antioxidants: Back and Forth in the Pathophysiology of Chronic Diseases, *Front Physiol.*, **11**, 694 (2020)
18. Narayanan Ravishankar, Shivaraj Chandrasekaran, Seenii Sooria Muthu, Joseph Jerrine and Raaman Nanjian, Antioxidant activities and phytochemical analysis of methanol extract of leaves of *Hypericum hookerianum*, *Int. J. Pharm. Pharm. Sci.*, **6**(4), 456-460 (2014)
19. Olatidoye, Paul Olawale, Ajani Olayemi Wasiu and Igwe U.N., Effect of Cooking Methods on Antioxidant Activity, Nitric Oxide Scavenging Activity and Phenolic Contents of Groundnuts Varieties Grown in Nigeria, *Journal of Agricultural Studies*, **4**(1), 2617-3530 (2020)
20. Orsavová J., Hlaváčková I., Mlček J., Snopek L. and Mišurcová L., Contribution of phenolic compounds, ascorbic acid and vitamin E to antioxidant activity of currant (*Ribes L.*) and gooseberry (*Ribes uva-crispa L.*) fruits, *Food Chemistry*, **284**, 323-333 (2019)
21. Prakash Om, Baskaran Revathy and Kudachikar V.B., Characterization, quantification of free, esterified and bound phenolics in Kainth (*Pyrus pashia* Buch-Ham. Ex D.Don) fruit pulp by UPLC-ESI-HRMS/ MS and evaluation of their antioxidant activity, *Food Chemistry*, **299**, 125114 (2019)
22. Raharjo Danang Haryoto, Antioxidant Activity of Mangrove *Sonneratia caseolaris L* using the FRAP Method, *International Summit on Science Technology and Humanity*, 623-629(2019)
23. Reuter S., Gupta S.C., Chaturvedi M.M. and Aggarwal B.B., Oxidative stress, inflammation and cancer: how are they linked?, *Free Radic Biol Med.*, **149**(11), 1603-16 (2010)
24. Smetanska Iryana, Alharthi Salman S. and Selim Khaled A., Physicochemical, antioxidant capacity and color analysis of six honeys from different origins, *Journal of King Saud University – Science*, **33**(5), 101447 (2021)
25. Widjaja Gunawan, Mukhamdov Bakhrambek, Mahmudiono Trias, Shalaby Mohammed Nader, Shichiyakh Rustem Adamovich, Rudiansyah Mohammad, Hasan Mohammed Khudair, Al-Dhalimy Aiman Mohammed Baqir and Jalil Abduladheem Turki, Evaluating the antioxidant activity in different citrus genotypes, *Food Science and Technology*, **43**, 1-7 (2023).

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